

Materials Required:

- 1 towelette
- 1 urine CCM vacuum tube
- 1 specimen vacuum collection cup
- 1 biohazard bag with absorbent material
- 2 Barcodes with DOB



Clean Catch Specimen Collection

To ensure safety and validity of the sample it is important to follow the instructions provided below.

Standard Procedure

Patient

1. Wash hands with soap and warm water and then rinse and dry hands.
2. Open the specimen collection cup by unscrewing the lid and setting the lid aside face DOWN. DO NOT touch the inside of the collection cup.
3. Follow gender specific instructions:

Females:

Spread the labia (folds of skin) apart with one hand and wipe with a towelette. Wipe from front to back. Repeat this process a second time with a fresh towelette.

Continue holding the labia apart. As you start to urinate, allow a small amount of urine to fall into the toilet bowl.

Males:

If uncircumcised, retract the foreskin.

Wipe the end of the penis with a towelette. As you start to

urinate, allow a small amount of urine to fall into the toilet bowl.

4. After the urine stream is well established, pass the collection cup into the urine stream and, after the cup is $\frac{1}{2}$ full, remove the cup from the urine stream and finish urinating into the toilet bowl.
5. Screw the lid on the cup tightly (DO NOT touch inside of cup or lid). Give the sealed cup to the nurse or attendant.

Nurse/Attendant

6. Remove the sticker from the lid of the cup, exposing the needle housed within the lid. Clean the gray stopper of the tube with an alcohol prep pad. Place the vacuum tube onto the exposed needle. The urine specimen will automatically fill the vacuum tube.
7. Remove tube from the lid and press sticker back into place
8. Place a completed barcode label on both the specimen and requisition. Place both into the same biohazard bag, seal and store at 2-8°C up to 72 hours (preferred) or room temp up to 24 hours.

Catheter Specimen Collection

Catheterized Patient Procedure

Specimen collection from patients with indwelling catheters requires aseptic technique.

DO NOT use lidocaine gel to numb the patient. Lidocaine gel can render a sample invalid or give false negative results.

1. Clamp off catheter tubing above the port to allow collection of freshly voided urine (minimum 2 ml urine required).
2. Vigorously clean the catheter port or wall of the tubing with 70% ethanol, and aspirate urine via either sterile needle (direct tubing puncture and aspiration), or syringe (if port has a Luer lock type fitting).
3. Eject the aspirated urine into the provided sterile specimen cup. Never submit urine for culture from the catheter collection bag.
4. Remove the sticker from the lid of the cup, exposing the needle housed within the lid. Place the vacuum tube onto the exposed needle. The urine specimen will automatically fill the vacuum tube.
5. Place a completed barcode label on both the specimen and requisition. Place both into the same biohazard bag, seal and store at 2-8°C up to 72 hours (preferred) or room temp up to 24 hours.

Specimen Stability & Transport

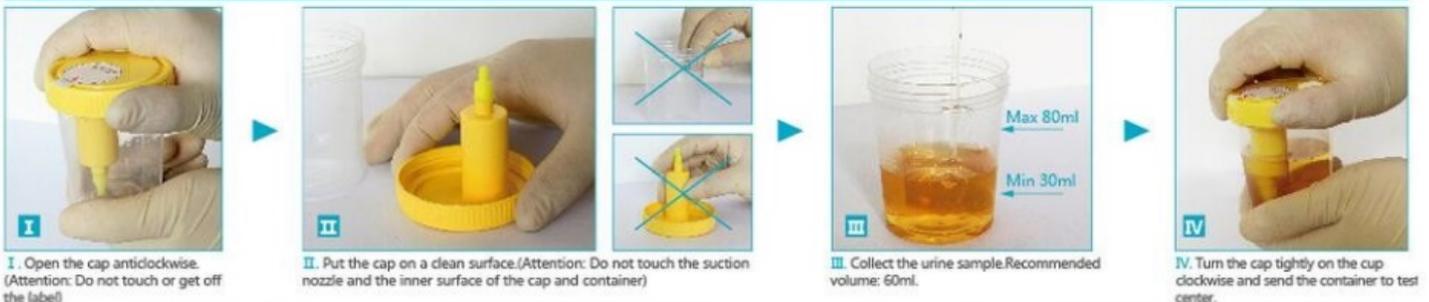
- Transport urine to the laboratory as soon as possible after collection.
- Specimens should be stored at 2-8°C up to 72 hours (preferred) or room temp for up to 24 hours. Specimens are stable at 2-8°C up to 7 days after collection, once received in the laboratory.
- Place completed requisition with matching barcode in the back pocket of the biohazard bag.
- Ship specimens in sealed biohazard bag in the provided clinical pak via UPS/FedEx via priority overnight shipping.

Important Considerations

- NEVER COLLECT URINE FROM A BEDPAN OR URINAL.
- Never submit urine for culture from the catheter collection bag.
- Soap rather than disinfectant is recommended for cleaning the urethral and/or vaginal area. If disinfectant is introduced into the urine during collection, it may be inhibitory to the growth of microorganisms.
- Lidocaine gel can render a sample invalid or give false negative results.
- The first morning voided urine is the best specimen due to the increased bacterial count after overnight incubation in the bladder.
- Be advised that forcing fluids to help the patient void will dilute the urine and may decrease the colony count. It is not recommended.
- If the patient is responsible for collection of specimen, he/she should be given specific, detailed instructions on collection procedure. Emphasize the importance of not contaminating the interior of the collection container by touching with hands or penis.
- PCR based test results will NOT be affected by antibiotic use.
- Stop taking probiotics and other supplements 2 days prior. Supplements that contain an active metabolite may degrade the DNA within your sample.
- Growth based sensitivity test results CAN be affected by antibiotic usage. Stop taking antibiotics 2 days prior. It's not always feasible to cease taking medications, and testing may still be possible while on antibiotics.

Instruction for use:

Urine collection process



Urine collection of medical staff:

